

Inheritance of purple pericarp colour in maize (*Zea mays* L.)

Salami Talhat Kolawole^a, Timilehin Gbemisola Oladipo^a, Ayodeji Abe^{b,*}

^aPan African University Life and Earth Sciences Institute (Including Health and Agriculture), University of Ibadan, Ibadan, Nigeria

^bDepartment of Crop and Horticultural Sciences, Faculty of Agriculture, University of Ibadan, Ibadan, Nigeria

*Corresponding Author: email: ayodabe@yahoo.com; Tel : +2348065019295

Abstract

Purple maize has high content of beneficial anthocyanin. Kernel colours and their intensity are associated with the concentration of phytochemicals and minerals, which could benefit human health. Breeding for purple-kernelled maize requires an understanding of its mode of inheritance. This study was conducted to investigate the mode of inheritance of purple pericarp colour in maize. A cross and its reciprocal was carried out between two maize lines: UIp01 with purple kernels and TZi3 which has white kernels. Ears bearing kernels of the six basic generations (P₁, P₂, F₁, F₂, BC₁P₁, and BC₁P₂) and F₃ derived from UIp01 × TZi3 (F₁) and TZi3 × UIp01 (RF₁) were sorted by colour, and grouped into purple and non-purple. Data were summarized for the distribution of kernel colour. Chi-square analysis with 95% confidence interval was carried out on the segregation ratio. The F₁ and RF₁ differed in pericarp colour. The F₁ had the purple maternal phenotype for pericarp colour, while the RF₁ had kernels with yellow pericarp. Irrespective of the direction of the cross, the F₂ segregated in the ratio 9:7 purple to non-purple, for complementary gene action. Segregation ratio of purple-kernelled to non-purple-kernelled ears in the F₃ had a good fit with ratio 1:3. Maternal effect was present in the inheritance of purple kernel colour in maize, with a complementary gene action in which the purple kernel colour was not completely dominant over the transparent phenotype, but the expression of purple kernel colour determined by the genotype of the parents.

Keywords: Complementary gene action, Inheritance pattern, Kernel colour, Maternal effect, Purple maize.

Introduction

Maize (*Zea mays* L.) is a cereal crop well adapted to a wide range of environments with a global total production of 1.24 billion tonnes with an average yield of 5.96 tonnes per hectare (FAOSTAT, 2023). It serves as an important food security crop in regions such as sub-Saharan Africa, Latin America and Southern Asia (Grote *et al.*, 2021; Soto-Gomez and Perez-Rodriguez, 2022; Bhushan *et al.*, 2024). In many parts of Africa, maize is an important staple food consumed in a wide variety of forms and as feed for animals. Apart from being rich in starch and protein, maize possesses the capacity to synthesize coloured pigments, particularly anthocyanins and carotenoids which are known to possess nutritional and functional roles (Nuss *et al.*, 2012; Rouf Shah *et al.*, 2016; Graham and Rosser, 2000; Bhushan *et al.*, 2024).

Maize has a vast genetic variation for kernel colour, which ranged from white through a succession of diverse shades of yellow, red, pink, purple, blue to black. However, the white and yellow variants are the most common commercial types (Ron-Parra *et al.*, 2016; Mendoza-Mendoza *et al.*, 2019). The diversity in maize types and the use to which it is put is closely associated with the culture of a given community and production system (Gomez *et al.*, 2000; Louette and Smale, 1998).

Purple maize is a flint maize type with its origin in the Andean region of South America and widely cultivated in countries like Peru, Mexico, Ecuador, Bolivia, Argentina, India and China, and distributed in the markets of Asia, United States of America and Europe (Colombo *et al.*, 2021; Kim *et al.*, 2023). Purple maize, like every other maize type, can be used as food, feed and raw materials for industries. However, it differs from the more common white and yellow maize due to its rich content of anthocyanins and phenolic compounds which are known to possess health-promoting effects (Lao and Giusti, 2016; Lao *et al.*, 2017; Wang *et al.*, 2024). These compounds are mainly located in pericarp or aleurone of purple maize (Bridle and Timberlake, 1997; Colombo *et al.*, 2021; Barba *et al.*, 2022). Studies have demonstrated that pigments in purple maize have anti-diabetic, anti-obesity, anti-hypertensive, anti-proliferative, anti-microbial, and anti-inflammatory properties (PenichePavía *et al.*, 2022; Kim *et al.*, 2023).

As a result of the distinct colour and antioxidant contents of purple maize, it is utilized as a natural functional food colorant in South America, Asia and Europe in substitution for unhealthy synthetic dyes (Moreno *et al.*, 2005; Jing *et al.*, 2007; Montilla *et al.*, 2011 and Lao *et al.*, 2017). Reports from previous studies (Cevallos-Casals and Cisneros-Zevallos, 2003; Wu *et al.*, 2006 and Li *et al.*, 2008) have shown that the average anthocyanin contents in purple maize kernels (16.4 mg g⁻¹) are considerably higher than those of known anthocyanin-rich plants like

blueberries (1.3 to 3.8 mg g⁻¹), strawberries (0.18 to 0.24 mg g⁻¹), red cabbage (2.81 to 3.63 mg g⁻¹), egg plants (8.57 mg g⁻¹) and chokeberries (14.8 mg g⁻¹). Despite the beneficial effects and utility of purple maize, there is limited awareness on its value in Africa. In Nigeria, purple maize is usually found as 'spots' in local farmers' maize fields. There are no known purple maize varieties in Nigeria, and limited research is devoted to its improvement and applications.

Elsewhere, studies are being conducted to increase the contents of phytochemicals in purple and red maize. Also, due to the association of the intensities of these colours with important phytochemicals and minerals, several efforts are being directed at transferring purple and red colours to adapted commercial varieties for the promotion of human health (Hong et al., 2020; Öztürk and Uzun, 2024). There is the need to develop and promote purple maize as a specialty corn with high contents of phenolic compounds and nutritional quality in Nigeria. Breeding for purple maize requires an understanding of its mode of inheritance and the genetic factors underlying the expression of the trait. A key component of such breeding goal is the investigation of patterns of inheritance of the purple pericarp trait. Outcomes from such studies could facilitate successful selection of parents, and the development of promising hybrids (Melchinger et al., 1986; Khamphan et al., 2018). Therefore, this study was conducted to investigate the mode of inheritance of purple pericarp colour in maize.

Materials and Methods

Experimental site

The study was conducted at the experimental field of the Department of Crop and Horticultural Sciences, Faculty of Agriculture, University of Ibadan (N07.45192°, E003.88994°, and 203 masl), Ibadan, Oyo State, Nigeria. The experimental field was cleared, and minimally tilled.

Genetic Materials, Crosses and Pericarp Colour Evaluation

Two maize inbred lines: TZ₁3 (formerly 1368) and UIp01 differing in kernel colour were used as genetic materials in this study. The line UIp01, a purple-kernelled flint maize inbred, was developed at the Department of Crop and Horticultural Sciences, University of Ibadan, Nigeria. The purple-kernelled maize was originally identified and collected from an open pollinated maize field and subjected to cycles of self-pollination and selection for purple kernels. Line TZ₁3, a white-kernelled flint maize inbred developed by the International Institute of Tropical Agriculture (IITA), Ibadan, is one of the parental lines of the commercial white-kernelled hybrid Oba Super-1.

The two maize lines were planted at different dates so as to synchronize flowering. The line UIp01 (P₁) was crossed as female parent to line TZ₁3 (P₂) to generate F₁. The reciprocal cross using TZ₁3 as female parent was also carried out to generate RF₁. Plants of the F₁ generations (including RF₁) were self-pollinated to produce F₂ kernels, and backcrossed to both parental lines to generate backcross populations. All pollinations were controlled and done by hand. Standard agronomic and field management practices for the cultivation of maize were followed. Harvesting was carried out ≥35 days after pollination when physiological maturity must be attained and air dried to about 12% moisture. Ears of the F₂ and backcross populations were harvested and processed individually. Ears of parental lines, F₁ and RF₁, F₂ and backcross populations were visually assessed for kernel colour, sorted by colour and counted. Thereafter, ears segregating for kernels were hand shelled, sorted by colour and counted to determine the inheritance pattern of the trait. The purple ears from the F₂ generations of both populations were then advanced to F₃. The ears were harvested at physiological maturity, processed individually, dried to 12% moisture content, assessed visually for kernel colour, sorted by kernel colour and counted.

Data Analysis

Data were summarized for the distribution of colours. Chi-square analysis was carried out on the segregation ratios of different kernel colours with a 95% confidence interval.

Results and Discussion

All the F₁ ears resulting from the cross between UIp01 (P₁) and TZ₁3 (P₂) were purple-kernelled, while the reciprocal cross (P₂ × P₁) yielded ears that were all yellow-kernelled (Figure 1). A comparison of the F₁ and RF₁ showed the presence of reciprocal difference in the phenotypic expression for purple pericarp kernel colour. The finding is consistent with previous reports in maize (Hossain *et al.*, 2019), and sesame (Laurentin and Benitez, 2014) on the existence of reciprocal differences. Differences in the pericarp phenotype of the F₁ and its reciprocal had been attributed to the influence of maternal genotype (Wilson and Hudson, 1979; Pandey *et al.*, 2013) in sesame, and Ron-Parra *et al.* (2016) in red pericarp grain of maize. The findings also show that nuclear genes may have no role in the inheritance of purple kernel colour in maize. However, in the present study, the purple and yellow pericarp colour of the F₁

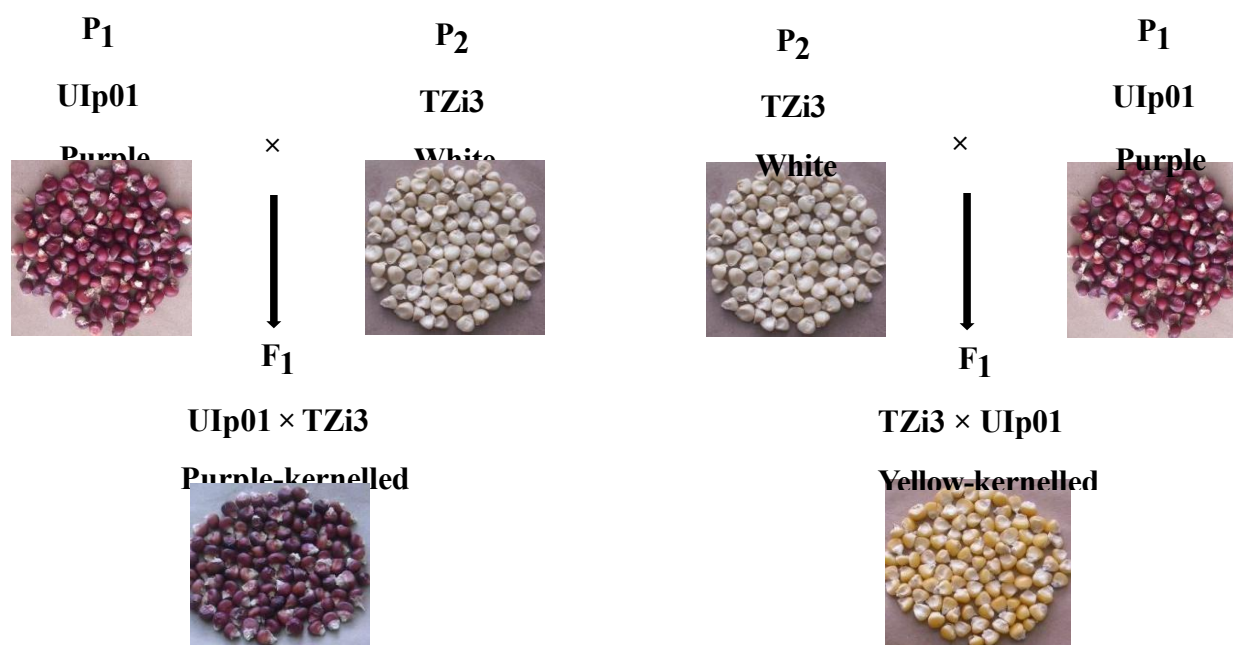


Figure 1. Distribution of kernel colours in the kernels of the F_1 hybrids arising from a cross between a purple (UIp01) and a white (TZi3) kernelled maize and RF_1 , respectively suggested that purple pericarp may not be dominant over transparent pericarp. Van-Deynze and Pauls (1994) had shown that the expression of seed coat colour in *Brassica*, though primarily controlled by the maternal genotype, is determined by the genetic constitution of the parents. The expression of yellow pericarp colour in the reciprocal cross in this study could be a consequence of complementary gene action. Our findings contradicts the results of Ron-Para *et al.* (2016) who had earlier reported maternal effect with complete dominance of the red phenotype over the transparent phenotype in the inheritance of red pericarp colour in maize.

Irrespective of the direction of the cross that generated the F_1 seeds, on selfing, the ears on the F_1 plants bearing F_2 kernels yielded cobs that were either all purple-kernelled or segregated for yellow and white kernels. No ear segregated for purple kernels on a cob (Figures 2 and 3). This shows that F_2 kernels on a cob do not segregate for purple pericarp. The findings from the present study however, contradicts the reports of Hossain *et al.* (2019) on the inheritance of red pericarp colour in maize, and Laurentin and Benitez (2014) on the inheritance of seed coat colour in sesame. The authors had reported that all the F_2 seeds independent of the direction of cross expressed the same phenotype. The proportion of purple kernelled ears to non-purple kernelled ears (ears that segregated for yellow and white kernels on a cob) had a good fit with 9:7 ratio (Table 1). This supports the digenic control of purple kernel colour in maize (Schoemaker *et al.*, 2024). Additionally, the segregation ratio of the ears segregating for yellow and white kernels on a cob was in conformity with the 3:1 ratio (Table 1). The direction of cross of the F_1 notwithstanding, backcrosses to the purple parental line yielded ears with purple kernels, while those to the white parental line yielded ears that segregated for yellow and white kernels (Table 1). The segregation of the yellow and white kernels arising from backcrosses to the white parental line had a good fit with the 1:1 ratio (Table 2).

On selfing the purple F_2 seeds derived from the different crosses, the ears on F_2 plants bearing F_3 kernels yielded four categories of ears. There were ears that were all purple-kernelled, those that segregated for yellow and white kernels on the cob, all yellow-kernelled or all white-kernelled. Similar to the observation for the F_2 seeds ears, no ear segregated for purple kernels on a cob. The ratio of purple-kernelled ears to non-purple kernelled ears (sum of ears that segregated for yellow and white kernels on a cob, all yellow-kernelled and all white-kernelled ears) had a good fit with 3:1 ratio expected for plant producing purple and non purple kernels (Table 2). Similar results were reported by Laurentin and Benitez (2014) for the inheritance of brown seed coat in sesame.

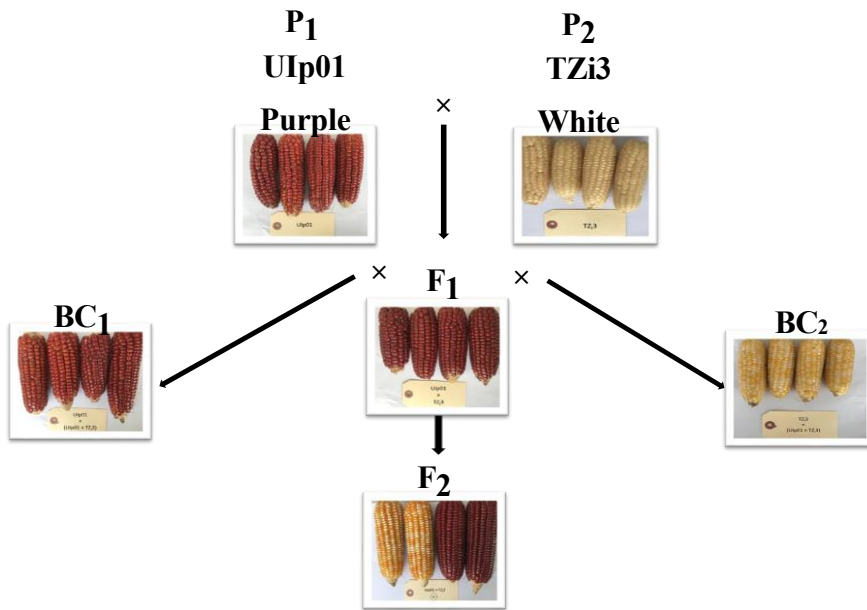


Figure 2. Distribution of kernel colour in the kernels of the F₂ and backcrosses of the cross UIp01 × TZi3

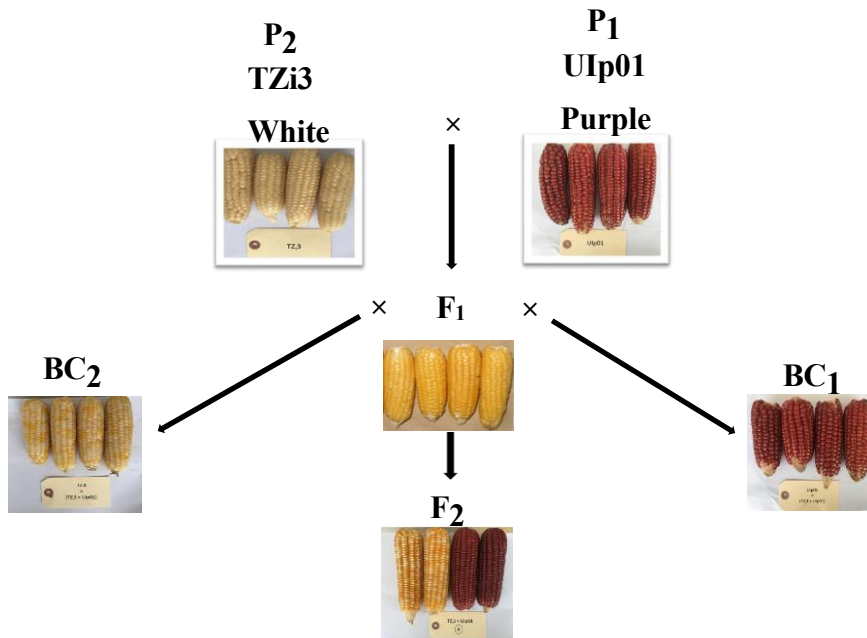


Figure 3: Distribution of kernel colour in the kernels of the F₂ and backcrosses of the cross TZi3 × UIp01

Table 1. Summary statistics of kernel phenotype and segregation ratios for purple pericarp in ears of the six basic generations and F₃ of reciprocal crosses between a purple maize line (UIp01) and a white maize line (TZi3) in Ibadan, Nigeria

Generation	Total number of ears	Number of ears with colour				Segregants	Ratio (Purple:non purple)	χ^2 calculate	χ^2 tabulated
		Purple	White	Yellow					
UIp01 (P ₁)	30	all							
TZi3 (P ₂)	30	all							
P ₁ × P ₂ (F ₁)	60	all							
P ₂ × P ₁ (RF ₁)	60	all							
F ₂	322	197			125	9:7	3.23	3.84	
RF ₂	320	175			145	9:7	0.32	3.84	
P ₁ × F ₁	68	all							
P ₂ × F ₁	88					all			
P ₁ × RF ₁	64	all							
P ₂ × RF ₁	111					all			
F ₃	300	230	13	14	43	3:1	0.44	3.84	
RF ₃	366	279	13	23	51	3:1	0.30	3.84	

Pp : Purple; W : White; Y : Yellow

Table 2: Segregation ratios of yellow to white on cobs of segregant ears of the F₂, RF₂, P₂ × F₁, P₂ × RF₁ generations of crosses between purple maize line UIp01 (P₁) and white maize line TZi3 (P₂)

Population	Number of segregant ears	Number of kernels per cob	Number kernels with colour		Ratio (Yellow:White)	χ^2	
			Yellow	White		calculated	tabulated
F ₂	125	417	309	108	3:1	0.205	3.84
RF ₂	145	408	302	106	3:1	0.209	3.84
P ₂ × F ₂	88	260	138	122	1:1	0.984	3.84
P ₂ × RF ₂	111	260	141	119	1:1	0.862	3.84

Conclusions and Recommendation

The inheritance of purple pericarp colour in maize was investigated by generating a cross and its reciprocal using a purple kernelled maize line Up01 and white kernelled maize line TZi3. Maternal effect was present in the inheritance of purple kernel colour in maize, with a complementary gene action in which the purple pericarp colour was not completely dominant over the transparent phenotype. The genetic constitution of the parental lines determined the expression of purple pericarp colour in maize. Our findings, which contradicts reports on the inheritance patterns of other kernel colours in maize, further demonstrates the considerable variability that exists in the expression of pericarp colours in maize. It is therefore important for the researcher to determine the inheritance pattern of the pericarp colour in the parents to be used.

Acknowledgements

This study benefited from the financial support provided by the African Union Commission through the Pan African University Institute of Life and Earth Sciences (PAULESI), University of Ibadan, Ibadan-Nigeria for the M.Sc, theses of the first and second authors, for which the authors are most grateful.

Conflict of interest

The authors declare no conflict of interest.

References

- Barba, F., Rajha, H.N., Debs, E., Abi-Khattar, A.M., Khabbaz, S., Dar, B.N., Simirgiotis, M., Castagnini, J.M., Maroun, R.G. and Louka, N. 2022. Optimization of Polyphenols' Recovery from Purple Corn Cobs Assisted by Infrared Technology and Use of Extracted Anthocyanins as a Natural Colorant in Pickled Turnip. *Molecules* 27: 5222.
- Bhushan, B., Kumar, S., Kaur, C., Devi, V., Chaudhary, D.P., Singh, A., Dagla, M.C., Karjagi, C.G., Saleena, L.A.K., Chandran, D., Kumar, M. 2024. Beyond colors: The health benefits of maize anthocyanins. *Applied Food Research* 4(1) : 100399. <https://doi.org/10.1016/j.afres.2024.100399>.
- Bridle, P. and Timberlake, C.F. 1997. Anthocyanins as natural food colours-selected aspects. *Food Chemistry* 58: 103 - 109.
- Cevallos-Casals, B.A. and Cisneros-Zevallos, L. 2003. Stoichiometric and kinetic studies of phenolic antioxidants from Andean purple corn and red-fleshed sweet potato. *Journal of Agriculture and Food Chemistry* 51: 3313 - 3319.
- Colombo, R., Ferron, L. and Papetti, A. 2021. Colored Corn: An Up-Date on Metabolites Extraction, Health Implication, and Potential Use. *Molecules* 26(1): 199. <https://doi.org/10.3390/molecules26010199>.
- FAOSTAT 2023. Food and Agriculture Organization of the United Nations statistical database. <https://www.fao.org/faostat/en/#data/QCL> (accessed on 19 May, 2025)
- Gomez, J.A.A., Bellon, M.R. and Smale, M. 2000. A regional analysis of maize biological diversity in Southeastern Guanajuato, Mexico. *Economic Botany* pp 60 - 72.
- Graham, R.D. and Rosser, J.M. 2000. Carotenoids in Staple Foods: Their Potential to Improve Human Nutrition. *Food and Nutrition Bulletin* 21(4): 404 - 409.
- Grote, U., Fasse, A., Nguyen, T.T. and Erenstein, O. 2021. Food security and the dynamics of wheat and maize value chains in Africa and Asia. *Frontiers in Sustainable Food Systems* 4: 617009. <https://doi.org/10.3389/fsufs.2020.617009>.
- Hong, H.T., Netzel, M.E. and O'hare, T.J. 2020. Anthocyanin composition and changes during kernel development in purple-pericarp supersweet sweetcorn. *Food chemistry* 315: 126284.
- Hossain, F., Bhat, S.R., Mohapatra, T. and Singh, A.K. 2019. Genetics on a maize cob: A teaching tool for schools. *Indian Journal of Genetics and Plant Breeding*, 79(1): Suppl. 340 - 366.

- Jing, P.U., Noriega, V., Schwartz, S.J. and Giusti, M.M. 2007. Effects of growing conditions on purple corn cob (*Zea mays* L.) anthocyanins. *Journal of Agricultural and Food Chemistry*. 55(21): 8625 - 8629.
- Khamphan, P., Lomthaisong, K., Harakotr, B., Ketthaisong, D., Scott, M.P., Lertrat, K. and Suriharn, B. 2018. Genotypic variation in anthocyanins, phenolic compounds and antioxidant activity in cob and husk of purple field corn. *Agronomy*. 8: 1 - 15.
- Kim, H.Y., Lee, K.Y., Kim, M. Hong, M., Deepa, P. and Kim, S. 2023. A Review of the Biological Properties of Purple Corn (*Zea mays* L.) *Scientia Pharmaceutica* 91(1): 6. <https://doi.org/10.3390/scipharm91010006>.
- Lao, F. and Giusti, M.M. 2016. Quantification of purple corn (*Zea mays* L.) anthocyanins using spectrophotometric and HPLC approaches: Method comparison and correlation. 9: 1367-1380.
- Lao, F., Sigurdson, G.T. and Giusti, M.M. 2017. Health benefits of purple corn (*Zea mays* L.) phenolic compounds. *Comprehensive Reviews in Food Science and Food Safety* 16: 234 - 246.
- Laurentin, H. and Benítez, T. 2014. Inheritance of seed coat color in sesame. *Pesquisa Agropecuária Brasileira* 49: 290 - 295.
- Li, F., Vallabhaneni, R. and Wurtzel, E.T. 2008. PSY3, a new member of the phytoene synthase gene family conserved in the Poaceae and regulator of abiotic-stress induced root carotenogenesis. *Plant Physiology* 146: 1333 - 1345.
- Louette, D. and Smale, M. 1998. Farmers' Seed Selection Practices and Maize Variety Characteristics in a Traditionally-Based Mexican Community. CIMMYT Economics Working Paper No. 98-04. Mexico, D.F.: CIMMYT.
- Melchinger, A.E., Geiger, H.H. and Schnell, F.W. 1986. Epistasis in maize (*Zea mays* L.) Genetic effects in crosses among early flint and dent inbred lines determined by three methods. *Theoretical and Applied Genetics*. 72: 231-239.
- Mendoza-Mendoza, C.G., Mendoza-Castillo, M.C., Castillo-González, F., Sánchez-Ramírez, F.J., Delgado-Alvarado, A., Pecina-Martínez, J.A. 2019. Agronomic Performance and Grain Yield of Mexican Purple Corn Populations from Ixtenco, Tlaxcala. *Maydica* (M21): 64-73.
- Montilla, E.C. Hillebrand, S., Antezana, A. and Winterhalter, P. 2011. Soluble and bound phenolic compounds in different Bolivian purple corn (*Zea mays* L.) cultivars. *Journal of Agricultural and Food Chemistry*. 53: 6649 - 6657.
- Moreno, Y.S., Sanches, G.S., Hernandez, D.R. and Lobato, N.R. 2005. Characterization of anthocyanin extracts from maize kernels. *Journal of Chromatographic Science* 43: 483 - 487.
- Nuss, E. T., Arscott, S. A., Bresnahan, K., Pixley, K. V., Rocheford, T., Hotz, C., Siasmusantu, W., Chileshe, J. and Tanumihardjo, S. A. 2012. Comparative intake of white-versus orange-colored maize by Zambian children in the context of promotion of biofortified maize. *Food and Nutrition Bulletin* 33(1): 63 - 71. <https://doi.org/10.1177/156482651203300106>.
- Öztürk, A. and Uzun, B. 2024. Development and morphological characterization of purple sweet corn lines. *Maydica* 67.
- Pandey, S.K., Das, A. and Dasgupta, T. 2013. Genetics of seed coat color in sesame (*Sesamum indicum* L.). *African Journal of Biotechnology* 12(42): 6061 - 6067.
- PenichePavía, H.A., Guzmán, T.J., MagañaCerino, J.M., GurrolaDíaz, C.M. and Tiessen, A. 2022. Maize flavonoid biosynthesis, regulation, and human health relevance: A Review. *Molecules*, 27(16): 5166. <https://doi.org/10.3390/molecules27165166>.

- Ron-Parra, J., Morales-Rivera, M.M., Jimenez-Lopez, J., Jimenez-Cordero, A.A., De La Cruz-Larios, L. and Sanchez-Gonzalez, J.J. 2016. Maternal genetic inheritance of red pericarp in the grain of maize. *Maydica*. 61-M21: 1-5.
- Rouf Shah, T., Prasad, K. and Kumar, P. 2016. Maize - A Potential Source of Human Nutrition and Health: A Review. *Cogent Food Agriculture* 2(1). <https://doi.org/10.1080/23311932.2016.1166995>.
- Schoemaker, D.L., Qiu, Y., de Leon, N., Hirsch, C.N. and Kaeppler, S.M. 2024. Genetic analysis of pericarp pigmentation variation in Corn Belt dent maize, *G3 Genes, Genomes, Genetics* 14(1): jkad256. <https://doi.org/10.1093/g3journal/jkad256>
- Soto-Gomez, D. and Perez-Rodriguez, P. 2022. Sustainable agriculture through perennial grains: Wheat, rice, maize, and other species. A review. *Agriculture, Ecosystem and Environment* 325: 107747. <https://doi.org/10.1016/j.agee.2021.107747>.
- Van-Deynze, A.E. and Pauls, K.P. 1994. Seed colour assessment in Brassica napus using a Near Infrared Reflectance spectrometer adapted for visible light measurements. *Euphytica* 76: 45 - 51.
- Wang, L., Yang, S., Yang, Y., Jiang, H., Huang, W., Bian, Y. and Li, B. 2024. Effects of endogenous anthocyanins from purple corn on the quality, physicochemical properties and antioxidant capacity of bread. *Journal of Food Measurement and Characterization* 18: 4678 - 4691. <https://doi.org/10.1007/s11694-024-02523-9>
- Wilson, V.E. and Hudson, L.W. 1979. Inheritance of lentil seedcoat mottle. *Journal of Heredity* 70(1): 83 - 84.
- Wu, X., Beecher, G.R., Holden, J.M., Haytowitz, D.B., Gebhardt, S.E. and Prior, R.L. 2006. Concentrations of anthocyanins in common foods in the United States and estimation of normal consumption. *Journal of Agricultural and Food Chemistry*. 54(11): 4069 -4075